

# Cerebral phaeohyphomycosis—a cure at what lengths?

Dong Ming Li, G Sybren de Hoog

Lancet Infect Dis 2009;  
9: 376–83

Peking University Third  
Hospital, Beijing, China  
(D M Li MD); and  
Centraalbureau voor  
Schimmelfcultures, Utrecht,  
Netherlands (D M Li,  
G S de Hoog PhD)

Correspondence to:  
Dong Ming Li, Peking university  
third hospital, Dermatology,  
No 49 North Huayuan Road,  
Beijing 100191, China  
dorisli@126.com

Cerebral phaeohyphomycosis is a fungal infection of the brain typically caused by *Cladophialophora bantiana*, *Exophiala dermatitidis*, and *Rhinoctadiella mackenziei*, all of which belong to the order Chaetothyriales. The disease results in black, necrotic brain tissue, black pus, and black cerebrospinal fluid. Pathogens usually reach the brain through the bloodstream or lymphatic fluid and occasionally through direct spreading or accidental inoculation. Patients can present with hemiparesis, tonic spasm, headache, fever, sensory variation, cerebral irritation, and even psychotic behavioural changes. Radiological images are characterised by ring-enhanced signs and hyperdense and hypodense lesions. Pathological features frequently include black-to-brown necrotic tissue or dark-coloured pus, granulomatous inflammation, giant cell vasculitis, and pigmented fungal elements, which are easily seen on a direct potassium hydroxide smear, a rapid method for diagnosis. Black fungi can be cultured from a biopsy specimen. Combined antifungal chemotherapy, surgical debridement, and careful immunological interventions are strongly recommended to eradicate these intractable infections.

## Introduction

Cerebral phaeohyphomycosis is the collective name for a group of cerebral fungal infections that are characterised by black necrotic tissue, black pus, and black cerebrospinal fluid (CSF).<sup>1–3</sup> If untreated, the infection leads to death within weeks, months, or, occasionally, years.<sup>1–6</sup> This infection is caused by a group of black fungi, and emerging pathogens from this group are still being encountered.<sup>5–12</sup> Most of the pathogens belong to a single order of environmental fungi, Chaetothyriales, according to new genetic studies.<sup>1</sup>

The sporadic and non-random distribution of these infections often leads them to be thought of as isolated events. Thus far, the infection has only been diagnosed after the pathogenic agent has been cultured from biopsy specimens, which delays treatment and results in high mortality.<sup>13–25</sup> Early diagnosis and effective therapy would improve patients' outcomes, but these depend largely on doctors' full understanding of the disease. This Review will focus on clinical signs and symptoms, radiographical features, pathology, diagnosis, and treatment, in addition to the aetiology, epidemiology, and risk factors of this infection.

## Aetiology and epidemiology

By the year 2000, 106 orders of fungi were recognised; 396 species in 25 orders were associated with infections in mammals, and more than 20 species were associated with systemic or disseminated human infection.<sup>1</sup> As already mentioned, most of the causal agents of cerebral phaeohyphomycosis belong to a single order of environmental fungi, Chaetothyriales, but a few are distributed in the orders Pleosporales, Sordariales, Xylariales, Helotiales, and Botryosphaerales (table). This group of fungi are melanised whether grown in vivo or in vitro, and they grow slowly and restrictedly.<sup>1</sup> Chaetothyriales includes several pathogens that can cause a disseminated fatal infection as well as a wide array of species that are involved in pulmonary, subcutaneous, and cutaneous infections. Three species of Chaetothyriales are primarily involved in brain infection: *Cladophialophora bantiana*, *Exophiala dermatitidis*, and *Rhinoctadiella mackenziei*.<sup>5,6,11,26–29</sup> Although emerging pathogens are still being identified (table),<sup>30–40</sup> this article focuses on the three primary species.<sup>7–12</sup>

*C bantiana*—previously known as *Cladosporium trichoides*, *Cladosporium bantianum*, and *Xylohypha emmonsii*—is the most frequently isolated pathogen from cases of cerebral phaeohyphomycosis; more than 70 cases of infection with this organism have been published.<sup>5,13,15–18,21–25,28,41</sup> The pathogen is distributed worldwide, although infections are most common in

	Order	Number of cases
<i>Neofusicoccum mangiferae</i>	Botryosphaerales	1
<i>Cladophialophora bantiana</i>	Chaetothyriales	>78
<i>Exophiala asiatica</i>	Chaetothyriales	1
<i>Exophiala dermatitidis</i>	Chaetothyriales	>25
<i>Exophiala jeanselmei</i>	Chaetothyriales	1
<i>Fonsecaea monophora</i>	Chaetothyriales	2
<i>Fonsecaea pedrosoi</i>	Chaetothyriales	1
<i>Rhinoctadiella mackenziei</i>	Chaetothyriales	21
<i>Neoscytalidium dimidiatum</i>	Helotiales	3
<i>Acrophialophora fusispora</i>	Incertae sedis	1
<i>Ochroconis gallopavum</i>	Incertae sedis	4
<i>Bipolaris hawaiiensis</i>	Pleosporales	2
<i>Bipolaris spicifera</i>	Pleosporales	2
<i>Bipolaris</i> sp	Pleosporales	1
<i>Curvularia lunata</i>	Pleosporales	4
<i>Curvularia pallescens</i>	Pleosporales	1
<i>Dissitimus exedrus</i>	Pleosporales	1
<i>Exserohilum rostratum</i>	Pleosporales	1
<i>Chaetomium atrobrunneum</i>	Sordariales	3
<i>Chaetomium globosum</i>	Sordariales	1
<i>Chaetomium perlucidum</i>	Sordariales	1
<i>Chaetomium strumarium</i>	Sordariales	2
<i>Phialemonium obovatum</i>	Sordariales	1
<i>Nodulisporium</i> sp	Xylariales	1

Table: Aetiological agents of cerebral phaeohyphomycosis

subtropical, non-arid climate zones.<sup>5,28</sup> Diagnosis depends largely on clinical disease recognition and pathogen identification.<sup>19,22,23,42,43</sup>

*E dermatitidis* has been known to cause brain infections in Japan since 1961, when it was called *Wangiella dermatitidis*.<sup>44</sup> Additional cases have been described from Japan, China, Korea, Pakistan, and Singapore.<sup>20,28,45–49</sup> Because most fatal brain infections were observed in young Asians with no history of immune disorder, *E dermatitidis* infection was previously assumed to be race-dependent.<sup>5</sup> However, more recent cases have included five cases of cerebral meningitis associated with steroid injections contaminated with *E dermatitidis* in the USA and 19 patients with *Exophiala jeanselmei* fungaemia (three of whom died because they did not receive therapy before culture results were known) associated with contaminated deionised water from a hospital pharmacy in Brazil. Consequently, cerebral phaeohyphomycosis is now believed to be a disease to which all people are susceptible.<sup>45,50–52</sup>

*R mackenziei*,<sup>29</sup> previously known as *Ramichloridium mackenziei* and *Ramichloridium schulzeri*, is a recently recognised endemic pathogen that causes cerebral phaeohyphomycosis.<sup>53–57</sup> Infections with this pathogen have all been reported in patients living in or originating from the Middle East—a hot, dry region. *R mackenziei* is the only neurotropic fungus that has been reported from this region.<sup>2,13,14,26,28,53–61</sup>

### Risk factors

Roughly half of patients with cerebral phaeohyphomycosis are otherwise healthy individuals, often in young men.<sup>5,6,45</sup> Infections have also been reported in patients receiving solid-organ transplants, with chronic organ failure, with primary or secondary immune-system disorders, with malignant diseases, with other infections, or patients who are on steroid therapy.<sup>6,16,62</sup> Among 54 patients with *C bantiana* infection, 21 were otherwise immunocompetent, five had an uncertain immune function, and ten cases were secondary to an infection with bacteria, parasites, or other fungi such as cryptococcus.<sup>5</sup> The remaining patients had chronic organ failure, had undergone organ transplantation or other surgery, or were on steroid therapy. In a review of 101 patients with cerebral fungal infections, 53 were immunocompetent and 29 had immune dysfunction (18 patients had received organ transplantations, ten had malignant diseases, and one had AIDS). Five patients were on steroid therapy, four were drug users, three had organ failure, one had diabetes, and six had trauma (one was trauma to the eye, two were trauma to the head, one was head surgery, one was from spinal acupuncture, and one was trauma at an unclear site).<sup>6</sup> Among 25 patients described from 1999 to 2004, infection was associated with contaminated medication in five, organ transplantation in five, chronic organ failure in three, malignant disorders in three, primary

immunodeficiency in one, AIDS in one, and chronic diseases in two; five had no prior disease.<sup>28,50</sup> Of the 20 patients that have been described since 2004, ten were immunocompetent, four had had organ transplantations, two had primary immunodeficiency, and four had chronic diseases.<sup>13–25</sup>

### Pathways to the brain

Chaetothyriales can reach the brain through blood or lymphatic vessels, by directly spreading from adjacent lesions, or by accidental inoculation.<sup>6,7,50</sup> Primarily, the fungi disseminate to the CNS through blood vessels; fungal elements have been frequently seen in arterial walls.<sup>17,23,24,43,63</sup> Because many patients have a pulmonary infection with the same agent before or at the same time as the cerebral infection, the potential for pulmonary entry has been hypothesised and confirmed in animals.<sup>1,52,55,64–65</sup> Exposure to soil or plant matter may be the source of respiratory infection.<sup>20,23,52</sup> The multiple brain abscesses that are commonly seen might also suggest dissemination through the bloodstream. Skin or pharyngeal trauma has also been suggested as a portal of entry.<sup>19,20,22,23,52,66</sup>

Secondarily, the fungus might reach the brain through lymphatic vessels. Cervical lymph node infection has often been reported before a cerebral infection that follows an event such as childbirth. Fungal elements have also been observed in lymphatic vessels.<sup>20,44,46</sup>

In some cases, the fungus infects the CNS by directly spreading from adjacent organs, such as the sinus and orbit.<sup>19,20</sup> Accidental inoculation has also been reported after an epidural injection with a contaminated steroid solution and through an open cranial wound.<sup>50,67,68</sup> These inoculations have allowed the mean incubation period to be calculated: about 1–3 months from inoculation to development of overt cerebral infection.<sup>50,67,68</sup>

Why the fungus has an affinity for brain tissues is unknown. An intriguing possibility is that melanin, which has been investigated as a toxic factor, may be responsible for CNS localisation.<sup>66,69,70</sup> Malignant melanoma has similarly high rates of CNS metastasis, more than 40% in an autopsy series.<sup>71</sup> Disrupting the polyketide synthase gene *WdPKS1* in *E dermatitidis* produces a melanin-deficient phenotype that is less virulent and more easily killed.<sup>70</sup>

### Clinical presentation

Clinical presentation of cerebral phaeohyphomycosis is typical of what would be expected with any brain lesion, including symptoms and signs such as seizures,<sup>17</sup> headache,<sup>17</sup> cerebral irritation, fever, and psychotic behavioural changes, although hemiparesis and hemisensory loss are the most common symptoms. Limb weakness is a common presenting symptom.<sup>2,13,17,21,60,65,72–77</sup> Unilateral hemisensory loss and paralysis may make cerebral phaeohyphomycosis seem more like a stroke or tumour, but because it is an infection, the disease usually

occurs concurrently with fever, headache, and other symptoms of infection. These symptoms are needed to differentiate the infection from other disorders with similar clinical manifestations. Symptoms like those of the common cold often appear at the beginning of infection, and as the disease progresses, other toxic symptoms might develop, such as fever with chills in the event of a comorbid bacterial infection.<sup>8,50,78</sup>

If the cranial nerve is involved, focal neurological deficits such as diplopia, vision loss, eyeball fixation, dysarthria, and ataxia would manifest.<sup>8,19,23,42,46,59,79</sup> Tonic spasm, chronic convulsion, atonic spasm, and focal motor seizures result from cerebral cell degeneration and necrosis.<sup>15,17,59,60,78,80–82</sup> Sometimes seizure is the first symptom, which can easily lead to misdiagnosis.<sup>17,82,83</sup>

Headache affects 55% of patients.<sup>6</sup> If headaches were caused by obstructive hydrocephalus, the pain would be extremely severe, similar to that of meningitis, but examination of CSF would show negative results, with the exception of expiratory changes in CSF pressure.<sup>42,46,48,75</sup>

Meningeal irritation could be a sign of cerebral phaeohyphomycosis, even in the absence of primary cerebrospinal meningeal infection. Most patients who had symptoms of meningitis, such as neck stiffness, did not show meningeal involvement in histopathology. The most likely reason for this finding is that brain tissue oedema increases the pressure of CSF, which causes meningeal signs.<sup>16,22,41</sup> Primary meningeal infection would most likely discolour CSF, or turn it black, and be accompanied by typical symptoms of headache, vomiting, stiffness, and papilloedema.<sup>15,20,47,50,55</sup> Chronic basal meningitis might result from mycotic aneurysm of the arteries.<sup>23</sup>

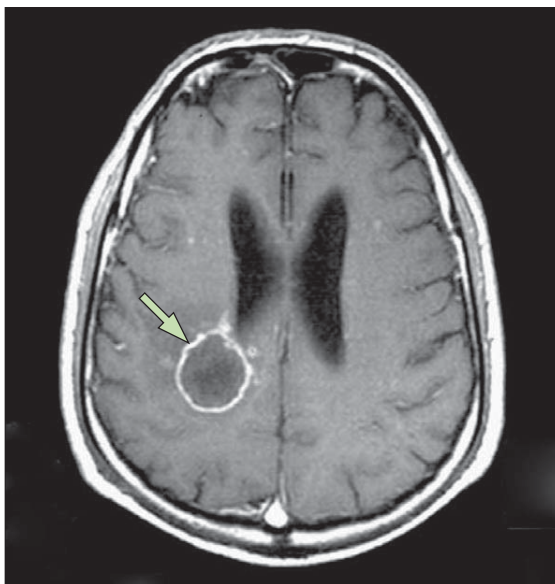


Figure 1: MRI showing ring-enhancing lesion in the right hemisphere of a brain infected with *Rhinocladiella mackenziei*  
Reproduced with permission from Elsevier Health.<sup>61</sup>

Lymphadenovariex (a varicose deformity of lymph nodes) with fever is another common sign before brain infection, primarily involving the cervical node. It appears months to years before the fungus moves into the brain, after which it spreads rapidly and is fatal in a matter of days.<sup>5,20,46</sup>

Concomitant symptoms depend on which other organs are involved. Respiratory infection with cough and sputum develops before or at the same time as brain infection in many patients.<sup>22,23,66</sup> Stomach ache, jaundice, hiccup, dark brown urine, liver dysfunction, and splenitis have all been recorded in patients with cerebral phaeohyphomycosis.<sup>61,79</sup>

CSF discolouration is a unique characteristic if the cerebrospinal membrane is involved; CSF appears black in *Exophiala* spp infection, green to brown in *R mackenziei* infection, and yellow-brown in *C bantiana* infection.<sup>20,42,43,47,50</sup> If the infection does not invade the cerebrospinal membrane, CSF test results are usually abnormal with hypoglycorrhachia, slight increases in protein concentration, and sometimes pleocytosis with eosinophils in addition to peripheral blood eosinophilia.<sup>43,48</sup>

### Radiological features

In most infections in immunocompetent patients, only a single lesion was seen; most patients with multiple lesions were immunocompromised.<sup>13–15,21,23,25,73,74,81,84–87</sup> In CT examinations, the typical finding is a ring-enhancing lesion with a low-attenuation core surrounded by hypodense lesions (figure 1).<sup>13,15,17,19,22,24,44,74,79,80,83,88,89</sup> MRI in *C bantiana* and *R mackenziei* infections reveals ring-enhancing lesions on T1-weighted images, hypointensity of the ring on T2-weighted images, and low-to-high signal intensity on diffusion-weighted imaging. The low attenuation core suggests the presence of necrotic tissue or pus, whereas the ring-enhancing sign usually represents a granulomatous lesion that is quite different from images of bacterial or parasitic infections, which typically manifest as multifocal, hypodense, non-enhancing lesions.<sup>83,88–90</sup>

In the early stages of infection, decreased attenuation can be seen in the highly attenuated zone of infected areas, which often leads to misdiagnosis as a brain tumour.<sup>23,48</sup> A typical ring-enhancing, hypodense lesion would appear as the infection progressed.<sup>8,21,44,48,60,78</sup> Irregular, variably contrast-enhancing masses can resemble either metastasis or high-grade glioma.<sup>42,43,91</sup>

### Pathological characteristics

On operation or autopsy, infected areas commonly appear, as with the CSF, as brown-to-black lesions in *Exophiala* spp infection, yellowish-brown in *C bantiana*, and dark greenish-brown in *R mackenziei*. Typically the abscess cavity has a thick wall filled with non-foul-smelling pus surrounded by hyperplastic tissue.<sup>8,54–56,79,92,93</sup>

Three major features might be observed under a microscope: granulomatous inflammation, dematiaceous



fungal material, and vasculitis with giant-cell infiltration. Granulomatous inflammation is the main histological feature, and it is usually characterised by the presence of multinucleated giant cells, histiocytes, lymphocytes, neutrophils, and necrotic debris with dematiaceous fungal materials (figure 2).<sup>8,9,15,17,26,46,63,74,75,94-97</sup>

Pigmented fungal elements must be found to conclusively diagnose cerebral phaeohyphomycosis. On a haematoxylin and eosin stain, these would be seen as dark-walled, branched, and septate hyphae, with possibly other forms of pseudohyphae, conidia, or budding cells, which would be stained intensely with Periodic acid-Schiff, Grocott's methenamine silver, or Congo red stain (figure 2).<sup>8,17,19,21,63,76,80,92,98,99</sup> Sometimes fungal elements can be seen inside the multinucleated giant cells.<sup>8,9,75,94</sup>

Vasculitis with giant-cell infiltration is an important characteristic in this infection, usually with mycotic thrombosis and subsequent artery infarction. Aneurysm occasionally occurs if the artery wall is destroyed.<sup>17,23</sup> Sometimes fungal stroke followed by mycotic thrombosis, aneurysm, or haemorrhage is the presenting sign that needs to be differentiated from a regular stroke.<sup>8,9,42,44-46,63,94,100</sup>

### Diagnosis

With typical clinical features and characteristic radiographical images, diagnosis is easy. However, pigmented hyphae and other fungal elements must be present to confirm the infection. Direct smear examination with potassium hydroxide is a rapid way to confirm the diagnosis, but examining a frozen section of the specimen is an alternative.<sup>101</sup> Black fungus can be recovered from the pus, brain-biopsy tissue, CSF, or blood when the specimen is inoculated on a culture medium like Sabouraud dextrose agar, malt extract agar, and potato dextrose agar and incubated at 27°C and 37°C.<sup>1</sup> Colonies vary in colour from dark green or brown to black.<sup>1</sup> Aetiological identity can be confirmed in reference laboratories.

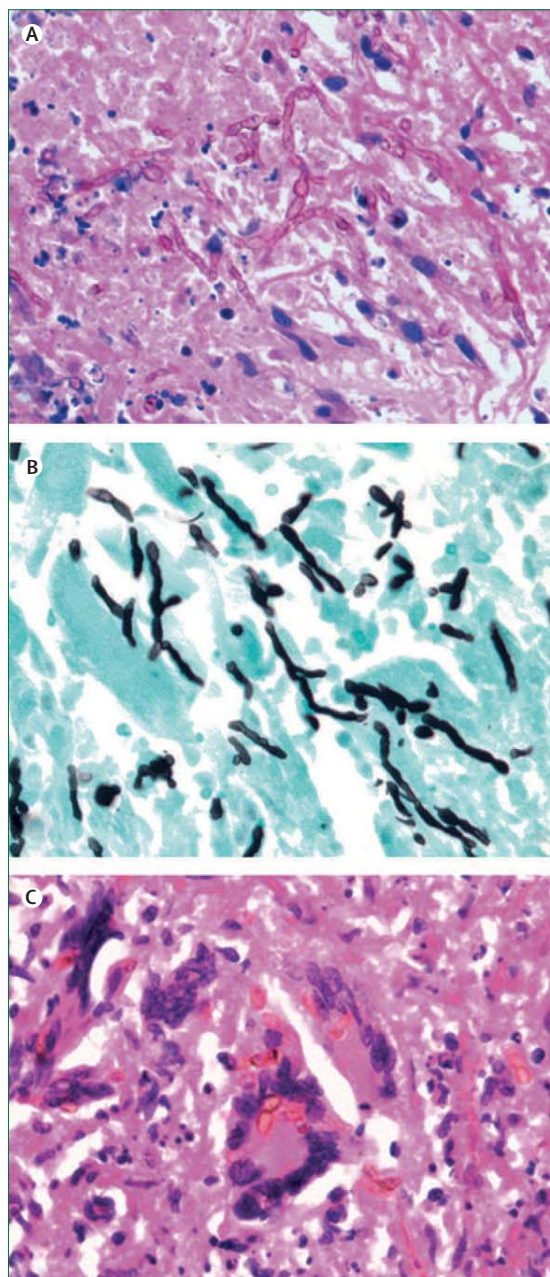
Although CT-guided stereotactic needle aspiration is a less invasive biopsy method, burr-hole tapping can be therapeutic (by draining pus) and provide biopsy specimens.<sup>2,21,25,74,102,103</sup> Lumbar puncture is helpful to examine CSF for factors that indicate meningitis if the cerebrospinal membrane is involved. Differential diagnoses include brain tumour, stroke, tuberculosis, and parasitic and other infections.<sup>42,58,93,104</sup>

### Treatment

Treatments include surgical debridement, combined antifungal therapy, and immune enhancement, although an isolated lesion can be removed successfully without chemotherapy.<sup>13,65,105,106</sup>

### Antifungal agents

Antifungal therapy should be combined with monitoring of antifungal susceptibility and clinical efficacy. Experiments on animals have shown that the in vivo



**Figure 2: Histopathological features of *Cladophialophora bantiana*** Pigmented fungal hyphae (A), elongated, and septate with unicellular lemon-shaped conidia (haematoxylin and eosin stain). The fungal elements (B) are intensely positive with Grocott's methenamine silver. Hyphae and conidia shown inside multinucleated giant cell (C), and in the surrounding brain parenchyma (hematoxylin and eosin stain). Magnification  $\times 220$ . Reproduced with permission from Wolters Kluwer Health.<sup>17</sup>

antifungal efficacy against *Cladophialophora bantiana* is consistent with in vitro antifungal susceptibility.<sup>13,107</sup> Three combinations are suggested: terbinafine plus an azole; an echinocandin (caspofungin, micafungin, and anidulafungin) plus amphotericin B or an azole; and flucytosine

with combinations of amphotericin B, itraconazole or posaconazole, or an echinocandin (these agents could enhance intracellular penetration and subsequent activity of flucytosine).<sup>13,83</sup> Nearly all successfully treated cases were treated with combination therapy;<sup>13,52</sup> monotherapy might seem effective initially, but it almost always results in treatment failure.<sup>2,3,8,11,13,21,50,52,60</sup> As new antifungal drugs like voriconazole and posaconazole become available, however, monotherapy may become more successful, because they have high efficacy, improved bioavailability, and are well tolerated.<sup>13,15,105,106,108–110</sup>

As high minimum inhibitory concentrations often result in treatment failure,<sup>19,107,110</sup> antifungal susceptibility should be monitored at the beginning and during the course of therapy because of the risk of resistance and cross-resistance induced by contact with antifungal drugs.<sup>108,111,112</sup> Alternating drugs might help avoid the development of resistance because long-term exposure to a particular regimen can induce resistance.<sup>80,113</sup> Regardless of in vitro susceptibility results, clinical and radiological improvement, which usually appears within a week or two of initiating treatment, is the best sign of clinical efficacy.<sup>13,86</sup>

Side-effects must be taken into account so that the treatment does not kill the patient before the fungus does.<sup>13,109</sup> Some antifungal agents, especially amphotericin B, are highly potent and can cause hypokalaemia, nephrotoxicity, and hepatotoxicity, any of which can be fatal.<sup>6,7,21,41</sup> However, long-term triazole therapy was reasonably safe in multiple studies and animal experiments.<sup>13</sup>

The duration of treatment necessary to eradicate the infection varies according to multiple factors, including the cause of disease, whether surgical intervention was necessary, and patient's immune status. Successful treatment courses have been as short as 3 months and lasted more than a year. Therapy should not be stopped until complete radiographical resolution occurs.<sup>15,21,76,81</sup>

### Surgical debridement

Surgical debridement of lesions is crucial in managing the infection. Debridement can greatly decrease the amount of fungal material at an infected site, and if the infection is small and localised, it can even be completely removed with a margin of adjacent, healthy tissue.<sup>17,24,52,114</sup> As brain abscesses form, pus must be drained because the thick wall around the abscess prevents drugs from penetrating. Burr-hole tapping and excision are surgical options.<sup>76</sup> Sometimes craniotomy must be done to excise a persistent abscess that resists antifungal drugs and repeated tapping. Meticulous care must be taken in the surgical procedure to prevent reinoculation of the fungus, which would lead to the recurrence and spread of the infection.<sup>15,19,52,59,83</sup>

### Immunotherapy

Immunological intervention might be needed for patients with intractable infections. For those who are not immunocompromised, interferons, interleukins, trans-

fused leucocytes, and colony-stimulating factors might enhance immunity by directly stimulating the immune system and might be useful as adjunct therapy.<sup>52,115–118</sup> In patients with compromised immune systems due to organ transplant, malignancy, or other factors, immune-suppressing drugs might need to be adjusted to fight fungal infections, carefully balancing aggressive antifungal treatment with the original therapy (antirejection drugs, chemotherapy).<sup>52,74</sup> Interferons, such as interferon  $\gamma$ , a protein produced in response to infections, might be a powerful weapon in the battle against fungal infections in immunocompromised patients.<sup>119</sup>

### Prognosis

Mortality for cerebral phaeohyphomycosis can be as high as 100% without treatment. For those who are treated with surgery and chemotherapy, mortality is still about 65%.<sup>13,14</sup> Matsumoto<sup>98</sup> reviewed 21 cases of systemic *Exophiala* spp infections and mortality was 48%, and the infection had a notable tendency to invade the CNS. In a review of ten culture-proven *C. bantiana* infections, mortality was 50% after patients underwent surgery that was followed by combination antifungal therapy. Of the 20 infections reported since 2004, only five patients survived.<sup>13–25,117–119</sup>

### Conclusions

Although cerebral phaeohyphomycosis has been recognised for decades, the infection is still poorly understood and is easily overlooked. The mortality rate of this disease remains high. Cerebral phaeohyphomycosis should be suspected in a patient who presents with hemiparesis, tonic spasm, or sensory variation, or all of these symptoms, and with symptoms of infection such as headache, fever, and fatigue. If a ring-enhancing lesion is seen on CT images, CT-guided stereotactic needle aspiration is strongly recommended to produce a biopsy specimen for potassium hydroxide examination, cultivation, and histopathology. Giant-cell vasculitis or granulomatous inflammation suggests the infection, and pigmented hyphae or conidia confirm the diagnosis. The aetiological agent can be identified by culture. Treatment

#### Search strategy and selection criteria

We searched Medline for English-language manuscripts limited to "human" and "case reports", "letters", "reviews" and "clinical conferences" from 1966 to 2008. We used MeSH terms "mycoses", "phaeohyphomycosis", or "chromoblastomycosis", in combination with MeSH term "brain" for brain fungal infections. The results were combined with "Exophiala" (MeSH), "Cladosporium" (MeSH) or its synonyms "Cladophialophora" or "Xylohypha", "Rhinochloidiella" or its synonym "Ramichloridium", respectively.

should begin as soon as possible and might require antifungal chemotherapy, surgical debridement, and immunological intervention. Cerebral phaeohyphomycosis can be cured with early diagnosis and aggressive therapy.

#### Conflicts of interest

We declare that we have no conflicts of interest.

#### Acknowledgments

We thank Ronnie Henry for revision of the paper and Teun Boekhout for scientific revision. This report was supported by National Natural Science Foundation of China (C30570003).

#### References

- De Hoog GS, Guarro J, Gene J, et al. Atlas of clinical fungi, 2nd edn. Utrecht: Centraalbureau voor Schimmelcultures, 2000.
- Podnos YD, Anastasio P, De La Maza L, et al. Cerebral phaeohyphomycosis caused by *Ramichloridium obovoideum* (*Ramichloridium mackenziei*): case report. *Neurosurgery* 1999; 45: 372–75.
- Silveira ER, Resende MA, Mariano VS, et al. Brain abscess caused by *Cladophialophora* (*Xylohypha*) *bantiana* in a renal transplant patient. *Transpl Infect Dis* 2003; 5: 104–07.
- Matsumoto T, Matsuda T, McGinnis MR, et al. Clinical and mycological spectra of *Wangiella dermatitidis* infections. *Mycoses* 1993; 36: 145–55.
- Horré R, De Hoog GS. Primary cerebral infections by melanized fungi: a review. *Stud Mycol* 1999; 43: 176–93.
- Revankar SG, Sutton DA, Rinaldi MG. Primary central nervous system phaeohyphomycosis: a review of 101 cases. *Clin Infect Dis* 2004; 38: 206–16.
- Biggs PJ, Allen RL, Powers JM, et al. Phaeohyphomycosis complicating compound skull fracture. *Surg Neurol* 1986; 25: 393–96.
- Carter E, Boudreaux C. Fatal cerebral phaeohyphomycosis due to *Curvularia lunata* in an immunocompetent patient. *J Clin Microbiol* 2004; 42: 5419–23.
- Hart AP, Sutton DA, McFeeley PJ, et al. Cerebral phaeohyphomycosis caused by a dematiaceous *Scopulariopsis* species. *Clin Neuropathol* 2001; 20: 224–28.
- Sides EH 3rd, Benson JD, Padhye AA. Phaeohyphomycotic brain abscess due to *Ochroconis gallopavum* in a patient with malignant lymphoma of a large cell type. *J Med Vet Mycol* 1991; 29: 317–22.
- Surash S, Tyagi A, De Hoog GS, et al. Cerebral phaeohyphomycosis caused by *Fonsecaea monophora*. *Med Mycol* 2005; 43: 465–72.
- Umabala P, Lakshmi V, Murthy AR, et al. Isolation of a *Umadulisporium* species from a case of cerebral phaeohyphomycosis. *J Clin Microbiol* 2001; 39: 4213–18.
- Al-Abdely HM, Alkhunaizi AM, Al-Tawfiq JA, et al. Successful therapy of cerebral phaeohyphomycosis due to *Ramichloridium mackenziei* with the new triazole posaconazole. *Med Mycol* 2005; 43: 91–95.
- Amr SS, Al-Tawfiq JA. Aspiration cytology of brain abscess from a fatal case of cerebral phaeohyphomycosis due to *Ramichloridium mackenziei*. *Diagn Cytopathol* 2007; 35: 695–99.
- Deb S, Khan AK, Debasish B, et al. Intracranial necrotizing granuloma caused by *Cladophialophora bantiana*. *Neurol India* 2005; 53: 335–36.
- Delfino D, De Hoog S, Polonelli L, et al. Survival of a neglected case of brain abscess caused by *Cladophialophora bantiana*. *Med Mycol* 2006; 44: 651–54.
- Harrison DK, Moser S, Palmer CA. Central nervous system infections in transplant recipients by *Cladophialophora bantiana*. *South Med J* 2008; 101: 292–96.
- Jayakeerthi SR, Dias M, Nagarathna S, et al. Brain abscess due to *Cladophialophora bantiana*. *Indian J Med Microbiol* 2004; 22: 193–95.
- Levin TP, Baty DE, Fekete T, et al. *Cladophialophora bantiana* brain abscess in a solid-organ transplant recipient: case report and review of the literature. *J Clin Microbiol* 2004; 42: 4374–78.
- Li DM, Li RY, de Hoog GS, et al. High fatality due to *Exophiala* infections in China. *Mycoses* (in press).
- Lyons MK, Blair JE, Leslie KO. Successful treatment with voriconazole of fungal cerebral abscess due to *Cladophialophora bantiana*. *Clin Neurol Neurosurg* 2005; 107: 532–34.
- Pardo F, Ferrer E, Romero PA, et al. Cerebral phaeohyphomycosis due to *Cladophialophora bantiana*. *Enferm Infecc Microbiol Clin* 2006; 24: 593–94.
- Roche M, Redmond RM, O'Neill S, et al. A case of multiple cerebral abscesses due to infection with *Cladophialophora bantiana*. *J Infect* 2005; 51: e285–88.
- Takei H, Goodman JC, Powell SZ. Cerebral phaeohyphomycosis caused by *Cladophialophora bantiana* and *Fonsecaea monophora*: report of three cases. *Clin Neuropathol* 2007; 26: 21–27.
- Tunuguntla A, Saad MM, Abdalla J, et al. Multiple brain abscesses caused by *Cladophialophora bantiana*: a challenging case. *Tenn Med* 2005; 98: 227–8, 235.
- Brandt ME, Warnock DW. Epidemiology, clinical manifestations, and therapy of infections caused by dematiaceous fungi. *J Chemother* 2003; 15 (suppl 2): 36–47.
- Chandramukhi A, Ramadevi MG, Shankar SK. Cerebral cladosporiosis—a neuropathological and microbiological study. *Clin Neurol Neurosurg* 1983; 85: 245–53.
- Kantarcioglu AS, de Hoog GS. Infections of the central nervous system by melanized fungi: a review of cases presented between 1999 and 2004. *Mycoses* 2004; 47: 4–13.
- Arzanlou M, Groenewald JZ, Gams W, et al. Phylogenetic and morphotaxonomic revision of *Ramichloridium* and allied genera. *Stud Mycol* 2007; 58: 57–59.
- Crous PW, Slippers B, Wingfield MJ, et al. Phylogenetic lineages in the Botryosphaeriaceae. *Stud Mycol* 2006; 55: 235–53.
- Geramishoar M, Zomorodian K, Zaini F, et al. First case of cerebral phaeohyphomycosis caused by *Nattrassia mangiferae* in Iran. *Jpn J Infect Dis* 2004; 57: 285–86.
- Li DM, Li RY, de Hoog GS, Wang DL. *Exophiala asiatica*, a new species from a fatal case in China. *Med Mycol* 2009; 47: 101–09.
- Alabaz D, Kibar F, Arikan S, et al. Systemic phaeohyphomycosis due to *Exophiala* (*Wangiella*) in an immunocompetent child. *Med Mycol* 2009; published online Jan 31. DOI:10.1080/13693780802715815.
- Surash S, Tyagi A, De Hoog GS, et al. Cerebral phaeohyphomycosis caused by *Fonsecaea monophora*. *Med Mycol* 2005; 43: 465–72.
- Ikram A, Hussain W, Satti ML, Wiqar MA. Invasive infection in a young immunocompetent soldier caused by *Scytalidium dimidiatum*. *J Coll Physicians Surg Pak* 2009; 19: 64–66.
- Aribandi M, Bazan III C, Rinaldi MG. Magnetic resonance imaging findings in fatal primary cerebral infection due to *Chaetomium strumarium*. *Australas Radiol* 2005; 49: 166–69.
- Mani RS, Chickabasaviah YT, Nagarathna S, et al. Cerebral phaeohyphomycosis caused by *Scytalidium dimidiatum*: a case report from India. *Med Mycol* 2008; 46: 705–11.
- Tan DH, Sigler L, Gibas CF, Fong IW. Disseminated fungal infection in a renal transplant recipient involving *Macrophomina phaseolina* and *Scytalidium dimidiatum*: case report and review of taxonomic changes among medically important members of the Botryosphaeriaceae. *Med Mycol* 2008; 46: 285–92.
- Al-Mohsen IZ, Sutton DA, Sigler L, et al. *Acrophialophora fusispora* brain abscess in a child with acute lymphoblastic leukemia: review of cases and taxonomy. *J Clin Microbiol* 2000; 38: 4569–76.
- Filizzola MJ, Martinez F, Rauf SJ. Phaeohyphomycosis of the central nervous system in immunocompetent hosts: report of a case and review of the literature. *Int J Infect Dis* 2003; 7: 282–86.
- Baddley JW, Salzman D, Pappas PG. Fungal brain abscess in transplant recipients: epidemiologic, microbiologic, and clinical features. *Clin Transplant* 2002; 16: 419–24.
- Gugnani HC, Sidiqqi N, Magulike MO. Cerebral phaeohyphomycosis: report of a case from Nigeria. *Mycoses* 1998; 41: 431–32.
- Emmens RK, Richardson D, Thomas W, et al. Necrotizing cerebritis in an allogeneic bone marrow transplant recipient due to *Cladophialophora bantiana*. *J Clin Microbiol* 1996; 34: 1330–32.
- Sasano N, Okamoto T, Takahashi T, et al. An autopsy case of primary chromoblastomycosis arising from the internal organs. Dark-brown granulomas in the liver and the brain without skin symptoms, observed in a smoking child 3 years old. *Tohoku J Exp Med* 1961; 73: 180–90.



- 45 Tintelnot K, de Hoog GS, Thomas E, et al. Cerebral phaeohyphomycosis caused by an *Exophiala* species. *Mycoses* 1991; **34**: 239–44.
- 46 Hiruma M, Kawada A, Ohata H, et al. Systemic phaeohyphomycosis caused by *Exophiala dermatitidis*. *Mycoses* 1993; **36**: 1–7.
- 47 Ajanee N, Alam M, Holmberg K, Khan J. Brain abscess caused by *Wangiella dermatitidis*: case report. *Clin Infect Dis* 1996; **23**: 197–98.
- 48 Chang CL, Kim DS, Park DJ, et al. Acute cerebral phaeohyphomycosis due to *Wangiella dermatitidis* accompanied by cerebrospinal fluid eosinophilia. *J Clin Microbiol* 2000; **38**: 1965–66.
- 49 De Hoog GS, Queiroz-Telles F, Haase G, et al. Black fungi: clinical and pathogenic approaches. *Med Mycol* 2000; **38** (suppl 1): 243–50.
- 50 Centers for Disease Control and Prevention. *Exophiala* infection from contaminated injectable steroids prepared by a compounding pharmacy—United States. *JAMA* 2003; **289**: 291–93.
- 51 Nucci M, Akiti T, Barreiros G, et al. Nosocomial fungemia due to *Exophiala jeanselmei* var. *jeanselmei* and a *Rhinocladiella* species: newly described causes of bloodstream infection. *J Clin Microbiol* 2001; **39**: 514–18.
- 52 Kenney RT, Kwon-Chung KJ, Waytes AT, et al. Successful treatment of systemic *Exophiala dermatitidis* infection in a patient with chronic granulomatous disease. *Clin Infect Dis* 1992; **14**: 235–42.
- 53 Rippon JW, Arnow PM, Larson RA, et al. 'Golden tongue' syndrome caused by *Ramichloridium schulzeri*. *Arch Dermatol* 1985; **121**: 892–94.
- 54 Campbell CK, Al-Hedaithy SSA. Phaeohyphomycosis of the brain caused by *Ramichloridium mackenziei* sp nov in Middle Eastern countries. *J Med Vet Mycol* 1993; **31**: 325–32.
- 55 Naim-Ur-Rahman, el Sheikh Mahgoub, Abu Aisha H, et al. Cerebral phaeohyphomycosis. *Bull Soc Pathol Exot Filiales* 1987; **80**: 320–28.
- 56 Naim-ur-Rahman, Mahgoub ES, Chagla AH. Fatal brain abscesses caused by *Ramichloridium obovoideum*: report of three cases. *Acta Neurochir (Wien)* 1988; **93**: 92–95.
- 57 del Palacio-Hernanz A, Moore MK, Campbell CK, et al. Infection of the central nervous system by *Rhinocladiella atrovirens* in a patient with acquired immunodeficiency syndrome. *J Med Vet Mycol* 1989; **27**: 127–30.
- 58 Kanj SS, Amr SS, Roberts GD. *Ramichloridium mackenziei* brain abscess: report of two cases and review of the literature. *Med Mycol* 2001; **39**: 97–102.
- 59 Khan ZU, Lamdhade SJ, Johnny M, et al. Additional case of *Ramichloridium mackenziei* cerebral phaeohyphomycosis from the Middle East. *Med Mycol* 2002; **40**: 429–33.
- 60 Sutton DA, Slifkin M, Yakulis R, et al. U.S. case report of cerebral phaeohyphomycosis caused by *Ramichloridium obovoideum* (*R mackenziei*): criteria for identification, therapy, and review of other known dematiaceous neurotropic taxa. *J Clin Microbiol* 1998; **36**: 708–15.
- 61 Alkhunaizi AM, Amir AA, Al-Tawfiq JA. Invasive fungal infections in living unrelated renal transplantation. *Transplant Proc* 2005; **37**: 3034–37.
- 62 Walz R, Bianchin M, Chaves ML, et al. Cerebral phaeohyphomycosis caused by *Cladophialophora bantiana* in a Brazilian drug abuser. *J Med Vet Mycol* 1997; **35**: 427–31.
- 63 Küllü S, Onol B, Kuştimur S, et al. Cerebral cladosporiosis. *Surg Neurol* 1985; **24**: 437–40.
- 64 Brenner SA, Morgan J, Rickert PD, Rimland D. *Cladophialophora bantiana* isolated from an AIDS patient with pulmonary infiltrates. *J Med Vet Mycol* 1996; **34**: 427–29.
- 65 Osiyemi OO, Dowdy LM, Mallon SM, et al. Cerebral phaeohyphomycosis due to a novel species: report of a case and review of the literature. *Transplantation* 2001; **71**: 1343–46.
- 66 Dixon DM, Polak A, Szanislo PJ. Pathogenicity and virulence of wild-type and melanin-deficient *Wangiella dermatitidis*. *J Med Vet Mycol* 1987; **25**: 97–106.
- 67 Barron MA, Sutton DA, Veve R, et al. Invasive mycotic infections caused by *Chaetomium perlucidum*, a new agent of cerebral phaeohyphomycosis. *J Clin Microbiol* 2003; **41**: 5302–07.
- 68 Kim RC, Hodge CJ Jr, Lamberson HV Jr, et al. Traumatic intracerebral implantation of *Cladosporium trichoides*. *Neurology* 1981; **31**: 1145–48.
- 69 Casadevall A, Rosas AL, Nosanchuk JD. Melanin and virulence in *Cryptococcus neoformans*. *Curr Opin Microbiol* 2000; **3**: 354–58.
- 70 Feng B, Wang X, Hauser M, et al. Molecular cloning and characterization of WdPKS1, a gene involved in dihydroxynaphthalene melanin biosynthesis and virulence in *Wangiella (Exophiala) dermatitidis*. *Infect Immun* 2001; **69**: 1781–94.
- 71 Bafaloukos D, Gogas H. The treatment of brain metastases in melanoma patients. *Cancer Treat Rev* 2004; **30**: 515–20.
- 72 Aldape KD, Fox HS, Roberts JP, et al. *Cladosporium trichoides* cerebral phaeohyphomycosis in a liver transplant recipient. Report of a case. *Am J Clin Pathol* 1991; **95**: 499–502.
- 73 Gupta SK, Manjunath-Prasad KS, Sharma BS, et al. Brain abscess in renal transplant recipients: report of three cases. *Surg Neurol* 1997; **48**: 284–87.
- 74 Salama AD, Rogers T, Lord GM, Lechler RI, Mason PD. Multiple *Cladosporium* brain abscesses in a renal transplant patient: aggressive management improves outcome. *Transplantation* 1997; **63**: 160–62.
- 75 Sujit Kumar GS, Dugar M, Chacko G. Cerebral phaeohyphomycosis presenting as an intraventricular mass. *Neurol India* 2006; **54**: 102–03.
- 76 Vyas MC, Joshi YR, Bhargava N, et al. Cerebral chromoblastomycosis—a rare case report of cerebral abscess and brief review of literature—a case report. *Indian J Pathol Microbiol* 2000; **43**: 81–85.
- 77 Dixon DM, Walsh TJ, Merz WG, et al. Infections due to *Xylohypha bantiana* (*Cladosporium trichoides*). *Rev Infect Dis* 1989; **11**: 515–25.
- 78 Kashgari TQ, Al-Miniawi H, Moawad Hanna MK. Cerebral phaeohyphomycosis caused by *Ramichloridium mackenziei* in the Eastern Province of Saudi Arabia. *Ann Saudi Med* 2000; **20**: 457–60.
- 79 Keyser A, Schmid FX, Linde HJ, et al. Disseminated *Cladophialophora bantiana* infection in a heart transplant recipient. *J Heart Lung Transplant* 2002; **21**: 503–05.
- 80 Lee YM, Tambyah PA, Lee KH, et al. Successful treatment of *Xylohypha bantiana* brain abscess mimicking invasive cerebral aspergillosis in a liver transplant recipient. *J Infect* 2003; **47**: 348–51.
- 81 Sood P, Dogra V, Thakur A, Mishra B, et al. Brain abscess due to *Xylohypha bantiana*. *Scand J Infect Dis* 2000; **32**: 708–09.
- 82 Saberi H, Kashfi A, Hamidi S, et al. Cerebral phaeohyphomycosis masquerading as a parafalcian mass: case report. *Surg Neurol* 2003; **60**: 354–59.
- 83 Lirmg JF, Tien RD, Osumi AK, et al. Cerebral phaeohyphomycosis complicated with brain abscess: a case report. *Zhonghua Yi Xue Za Zhi (Taipei)* 1995; **55**: 491–95.
- 84 Türker A, Altınörs N, Aciduman A, et al. MRI findings and encouraging fluconazole treatment results of intracranial *Cladosporium trichoides* infection. *Infection* 1995; **23**: 60–62.
- 85 Buxi TB, Prakash K, Vohra R, et al. Imaging in phaeohyphomycosis of the brain: case report. *Neuroradiology* 1996; **38**: 139–41.
- 86 Trinh JV, Steinbach WJ, Schell WA, et al. Cerebral phaeohyphomycosis in an immunodeficient child treated medically with combination antifungal therapy. *Med Mycol* 2003; **41**: 339–45.
- 87 Mukherji SK, Castillo M. Cerebral phaeohyphomycosis caused by *Xylohypha bantiana*: MR findings. *AJR Am J Roentgenol* 1995; **164**: 1304–05.
- 88 Whelan MA, Hilal SK. Computed tomography as a guide in the diagnosis and follow-up of brain abscesses. *Radiology* 1980; **135**: 663–71.
- 89 Britt RH, Enzmann DR. Clinical stages of human brain abscesses on serial CT scans after contrast infusion. Computerized tomographic, neuropathological, and clinical correlations. *J Neurosurg* 1983; **59**: 972–89.
- 90 Chapman ME, Sellar RJ, Whittle IR. Pyocephalus: a valuable CT finding in cerebral abscess. *Clin Radiol* 1992; **45**: 195–97.
- 91 Davis LE. Fungal infections of the central nervous system. *Neurol Clin* 1999; **17**: 761–81.
- 92 Salem FA, Kannangara DW, Nachum R. Cerebral chromomycosis. *Arch Neurol* 1983; **40**: 173–74.
- 93 Freitas A, Pedral-Sampaio DB, Espinheira Nogueira L, et al. *Cladophialophora bantiana* (previously *Cladosporium trichoides*): first report of a case in Brazil. *Braz J Infect Dis* 1997; **1**: 313–16.

- 94 Masini T, Riviera L, Capprici E, et al. Cerebral phaeohyphomycosis. *Clin Neuropathol* 1985; 4: 246–49.
- 95 Dastur HM, Chaukar AP, Rebello MD. Cerebral chromoblastomycosis due to *Cladosporium trichoides* (*Bantianum*), I: a review and case report. *Neurol India* 1966; 14: 1–5.
- 96 Fader RC, McGinnis MR. Infections caused by dematiaceous fungi: chromoblastomycosis and phaeohyphomycosis. *Infect Dis Clin North Am* 1988; 2: 925–38.
- 97 Heney C, Song E, Kellen A, et al. Cerebral phaeohyphomycosis caused by *Xylohypha bantiana*. *Eur J Clin Microbiol Infect Dis* 1989; 8: 984–88.
- 98 Matsumoto T, Matsuda T, McGinnis MR, et al. Clinical and mycological spectra of *Wangiella dermatitidis* infections. *Mycoses* 1993; 36: 145–55.
- 99 Palaoglu S, Sav A, Basak T, Yalcinlar Y, et al. Cerebral phaeohyphomycosis. *Neurosurg* 1993; 33: 894–97.
- 100 Moja M, Muthuphei MN, van der Westhuizen LR, et al. Multiple infarcts in a patient with cerebral phaeohyphomycosis: CT and MRI. *Neuroradiology* 2000; 42: 261–66.
- 101 Banerjee U, Mohapatra AK, Sarkar C, et al. Cladosporiosis (cerebral phaeohyphomycosis) of brain—a case report. *Mycopathologia* 1989; 105: 163–66.
- 102 Raut A, Muzumdar D, Narlawar R, Nagar A, Ahmed N, Hira P. Cerebral abscess caused by *Cladosporium bantianum* infection—case report. *Neurol Med Chir (Tokyo)* 2003; 43: 413–15.
- 103 Sekhon AS, Galbraith J, Mielke BW, et al. Cerebral phaeohyphomycosis caused by *Xylohypha bantiana*, with a review of the literature. *Eur J Epidemiol* 1992; 8: 387–90.
- 104 Nadkarni TD, Goel A, Shenoy A, et al. *Cladosporium bantianum* (trichoides) infection of the brain. *J Postgrad Med* 1993; 39: 43–44.
- 105 Gold WL, Vellend H, Salit IE, et al. Successful treatment of systemic and local infections due to *Exophiala* species. *Clin Infect Dis* 1994; 19: 339–41.
- 106 Schwartz S, Milatovic D, Thiel E. Successful treatment of cerebral aspergillosis with a novel triazole (voriconazole) in a patient with acute leukaemia. *Br J Haematol* 1997; 97: 663–65.
- 107 Li DM, Xiu DR, Li RY, Samson RA, de Hoog GS, Wang DL. *Aspergillus flavus* myositis in a patient after liver transplantation. *Clin Transplant* 2008; 22: 508–11.
- 108 Fica A, Diaz MC, Luppi M, et al. Unsuccessful treatment with voriconazole of a brain abscess due to *Cladophialophora bantiana*. *Scand J Infect Dis* 2003; 35: 892–93.
- 109 Pitisuttithum P, Negroni R, Graybill JR, et al. Activity of posaconazole in the treatment of central nervous system fungal infections. *J Antimicrob Chemother* 2005; 56: 745–55.
- 110 Garg N, Devi IB, Vajramani GV, et al. Central nervous system cladosporiosis: an account of ten culture-proven cases. *Neurol India* 2007; 55: 282–88.
- 111 Panackal AA, Gribskov JL, Staab JF, et al. Clinical significance of azole antifungal drug cross-resistance in *Candida glabrata*. *J Clin Microbiol* 2006; 44: 1740–43.
- 112 Middleton FG, Jurgenson PF, Utz JP, et al. Brain abscess caused by *Cladosporium trichoides*. *Arch Intern Med* 1976; 136: 444–48.
- 113 Elter T, Sieniawski M, Gossmann A, et al. Voriconazole brain tissue levels in rhinocerebral aspergillosis in a successfully treated young woman. *Int J Antimicrob Agents* 2006; 28: 262–65.
- 114 Sandhyamani S, Bhatia R, Mohapatra LN, et al. Cerebral cladosporiosis. *Surg Neurol* 1981; 15: 431–34.
- 115 Zhou Q, Murphy WJ. Immune response and immunotherapy to *Cryptococcus* infections. *Immunol Res* 2006; 35: 191–208.
- 116 Rachini A, Pietrella D, Lupo P, et al. An anti- $\beta$ -glucan monoclonal antibody inhibits growth and capsule formation of *Cryptococcus neoformans* in vitro and exerts therapeutic, anticryptococcal activity in vivo. *Infect Immun* 2007; 75: 5085–94.
- 117 Rossmann SN, Cernoch PL, Davis JR. Dematiaceous fungi are an increasing cause of human disease. *Clin Infect Dis* 1996; 22: 73–80.
- 118 Salaki JS, Louria DB, Chmel H. Fungal and yeast infections of the central nervous system. A clinical review. *Medicine (Baltimore)* 1984; 63: 108–32.
- 119 Kawakami K, Tohyama M, Teruya K, et al. Contribution of interferon- $\gamma$  in protecting mice during pulmonary and disseminated infection with *Cryptococcus neoformans*. *FEMS Immunol Med Microbiol* 1996; 13: 123–30.