

Phylogenetic analysis of *Alternaria* spp. associated with apple core rot and citrus black rot in South Africa

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Dry core rot of apple (DCR) and *Alternaria* black rot of citrus (AR) have in the past respectively been ascribed to *Alternaria alternata* and *A. citri*. In recent years, however, it has been speculated that several other species of *Alternaria* could also be associated with these diseases. In an attempt to elucidate the identity of these taxa, 25 isolates associated with DCR, and 26 isolates associated with AR were selected for molecular characterisation. Nucleotide sequences of 1116 sites including the histone gene section and the internal transcribed spacers ITS 1 and 2 of the rRNA gene were determined for these isolates. The gene trees generated from the individual and combined data sets using maximum parsimony, maximum likelihood and neighbour-joining analysis methods distinguished the clades with strong bootstrap support, namely *Alternaria* sp., *A. arborescens*, *A. infectoria*, *A. tenuissia*, and a clade containing isolates variable in morphology, referred to as the *Alternaria* group. In the alignment of the combined ITS and histone data set, unique transition/transversion substitutions, as well as positional insertions and deletions were observed for each of the above clades. In addition, key sequences in the form of serially coposing nucleotides in both the ITS and histone sections of the alignment were also discovered for the molecular identification of *A. arborescens*, *A. infectoria* and *A. tenuissia*. The final phylogeny also indicated that no host specificity existed among the species associated with these two post-harvest disease complexes. Contrary to the host specificity observed on leaf diseases of these hosts in the field, it appears that the post-harvest diseases are the result of adverse storage conditions and opportunism of different small-spored *Alternaria* spp.

INTRODUCTION

Core rot of apple and black rot of citrus are two well-known post-harvest diseases attributed to small-spored species of *Alternaria* (Serdani et al. 1998, Sions 1999a). Dry core rot (DCR) of apple, which mainly infects the Red Delicious varieties, has in the past been linked to a single species, *A. alternata* (Carpenter 1942, Miller 1959, Cobrinck et al. 1985b), while several *Alternaria* spp. have been associated with mouldy core of apples (Heald 1921, Ellis 1983). DCR is a worldwide problem, occurring in most countries where apples are grown (Serdani et al. 1998, including South Africa (Cobrinck et al. 1985a). Annual losses of between 6 and 8% are generally attributed to this disease (Cobrinck, van der Grinten & Grobbelaar 1984). A recent study on DCR that characterised isolates based on metabolite production, cultural characteristics, sporulation patterns, pathogenicity and DNA sequence analysis of the internal transcribed spacers ITS1 and 2 of the nuclear-encoded rRNA gene (rDNA), concluded that

representatives of several species-groups, including *A. arborescens*, *A. infectoria* and *A. tenuissia* were involved (Serdani et al. 2002). Results from inoculation studies suggested, however, that the *A. tenuissia* species-group was the major pathogen associated with DCR of Top Red apples in South Africa (Serdani et al. 2002). It has been reported in basidiomycetes, however, that DNA sequence data from the ITS region are insufficient to distinguish all species, as some taxa could evolve more quickly than mutations could accumulate in their ITS regions (Gardes et al. 1991, Anderson & Stasoski 1992). Furthermore, the same has also been found in some ascomycete genera, where sequence data of the β -tubulin and histone regions were required to distinguish species in the *Cylindrocladus pathiphylli* and *C. oridanus* species complexes that otherwise appeared similar based on their ITS data sets (Kang, Crous & Schoch 2001).

Ste-end rot (SER) or *Alternaria* black rot (AR) affects virtually all citrus varieties during prolonged periods of cold storage (Brown & McKornack 1972) and is also a serious problem in South Africa. Symptoms include ste-end browning as well as rot of the

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central axis (Shiffmann-Nadel et al. 1981). Various researchers have reported species of *Alternaria* to be associated with SER July 1967, Pelser La Grange 1981, Prusky, Fuchs auberan 1981 as well as with navel-end rot Schutte, eton Pelser 1994, of which *A. citri* has been identified as the main culprit (Brown & McKornack 1972, Shiffmann-Nadel et al. 1981). Although pathotypes of *A. alternata* have been linked to various diseases Kusaba Tsuge 1995, *A. citri* was originally described from internal dry rot symptoms of oranges Sions 1999a. Additional *Alternaria* diseases associated with citrus are *A. liicola* on Mexican lime *Citrus aurantiifolia* and other species of citrus, *Alternaria* brown spot on tangerine *C. reticulata* and related hybrids, and *Alternaria* leaf spot of rough lemon *C. abhiri* Sions 1999a. Sions 1990, 1999a found that the *Alternaria* spp. associated with citrus brown spot were neither *A. citri* nor *A. alternata*, but represented 10 new species of *Alternaria*, which were associated with these symptoms throughout different parts of the world. ABR, which is the citrus disease focused on in the present study, has traditionally been linked to *A. citri*. No ex-type cultures are available of *A. citri*, and its identity remains rather obscure. However, Sions 1999a stated that, based on the variation that he has encountered from AR isolates, species morphologically different from the type species of *A. citri* were also responsible for the disease. Based on sequence data of the ITS region, as well as RFLPs of tDNA, Kusaba Tsuge 1994, 1995, 1997) reported that there was little differentiation between host-specific toxin-producing citrus isolates, and isolates from hosts such as apple, strawberry and pear. The objectives of the present study were firstly to compare the *Alternaria* spp. causing DCR and AR using parsimony and likelihood analyses of multi-locus DNA sequence data of the histone and ITS regions, secondly to establish if any of the taxa were unique to either, or shared by both hosts, and lastly to determine if the histone data set could distinguish one species of *Alternaria* than delineated based on the ITS sequence data.

MATERIALS AND METHODS

Fungal cultures and growth conditions

Twenty-six *Alternaria* isolates from citrus species with AR symptoms 4 Navel oranges, 14 Minneola tangelo and 8 Cleontines were plated onto 9 cm Petri dishes containing potato carrot agar PCA 20 g potato agar, 20 g carrot, 20 g potato, 1000 l H₂O. Twenty-four *Alternaria* isolates obtained from a Top Red apple orchard with DCR, as well as one reference isolate of *A. infectoria* EGS 27-193 CR31 were also included and plated onto PCA Table 1. Plates were placed at 23 °C under an alternating 8/16 h cool-white fluorescent light/dark cycle Sions Roberts 1993 to induce

sporulation. A single spore from each isolate was transferred to Petri dishes containing PCA, and aseptically transferred to and stored on potato dextrose agar PDA 12 g potato agar, 200 g potatoes, 20 g sucrose, 1000 l H₂O slants at room temperature as well as in 20 l bottles containing sterile water for the purpose of long-term storage. All fungal cultures are held at the culture collection at the Department of Plant Pathology, University of Stellenbosch STE-U.

Application and sequencing

Genomic DNA was isolated from fungal mycelia collected from the plates using the isolation protocol of Lee Taylor 1990. Template DNA 20 ng was applied in a 25 µl PCR reaction mixture consisting of 10 mM KCl, 10 mM NH₄SO₄, 20 mM TrisHCl pH 8.8, 6 mM MgCl₂, and 500 µM each of dATP, dCTP, dGTP, and dTTP, with 60 pmol ITS1 and ITS4 primers White et al. 1990 or H3-1a and H3-1b Glass Donaldson 1995 primers, and 2.5 units Taq polymerase. The reaction was set up as follows: initial denaturation at 96 °C for 2 min, followed by 30 cycles of denaturation at 96 °C for 15 s, annealing at 55 °C for 30 s, extension at 75 °C for 35 s and final extension at 75 °C for 2 min in a RapidCycler Idaho Technology, Idaho. A negative control using water instead of template DNA was set up for each experiment. PCR products were analysed by electrophoresis at 75 °C for 2 h in a 0.8% agarose gel in 0.5 × TAE buffer (0.4 M Tris, 0.05 M NaAc, and 0.01 M EDTA, pH 7.85) and visualised under UV light in a transilluminator TF-35C, under Loura following ethidium bromide staining.

PCR products were purified by using a Quick PCR Purification Kit (Qiagen GmbH, Germany). The purified PCR products were sequenced using the ABI Prism 377 DNA Sequencer (Perkin-Elmer, Norwalk, CN). The cycle sequencing reaction with 2040 ng of DNA template and 3.2 pmol primer in a total volume of 10 µl was carried out with a Dye Terminator Cycle Sequencing Ready Reaction Kit containing AmpliTaq DNA Polymerase (Perkin-Elmer). The reaction was set up as 25 cycles of 96 °C for 30 s, 50 °C for 15 s, and 60 °C for 4 min. The resulting fragments were then purified using Centri-Sep Spin columns (Princeton Separations, Adelphia, NJ) and loaded onto the sequencing gel.

Phylogenetic analysis

Nucleotide sequences of the histone and ITS rDNA of this study Genbank accession nos AF404591 AF404641 and AF404642 AF404667 and those of the outgroup, *Fusarium* spp. Genbank nos 94174 and U34426 were assembled using Te-Edit Plus (Amersham Pharmacia Biotech, Little Chalfont, UK). The alignment of the sequences was conducted using the CLUSTAL W software (Thompson, Higgins & Gibson 1994). Adjustments for gaps were made by eye where

Table 1. Isolates of *Alternaria* spp. sapled froapple core rot and citrus black rot syptos in South Africa.

Accession no.	Other no.	Morpho-species ^a	Phylospecies	Genank accession no. ITS, Histone	ariety or cultiar ^b	Location ^c
CR 1	STE-U 4242	<i>A. arborescens</i>	<i>Alternaria</i> group, clade 1	AF397218, AF404591	A, Top Red	WC, Grabouw
CR 2	STE-U 4243	<i>A. arborescens</i>	<i>A. arborescens</i> , clade 3	AF397219, AF404592	A, Top Red	WC, Grabouw
CR 3	STE-U 4244	<i>A. arborescens</i>	<i>A. arborescens</i> , clade 3	AF397220, AF404593	A, Top Red	WC, Grabouw
CR 5	STE-U 4246	<i>A. arborescens</i>	<i>A. arborescens</i> , clade 3	AF397222, AF404594	A, Top Red	WC, Grabouw
CR 6	STE-U 4247	<i>A. arborescens</i>	<i>Alternaria</i> sp., clade 2	AF397223, AF404595	A, Top Red	WC, Grabouw
CR 7	STE-U 4248	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397224, AF404596	A, Top Red	WC, Grabouw
CR 8	STE-U 4249	<i>A. arborescens</i>	<i>Alternaria</i> group, clade 1	AF397225, AF404597	A, Top Red	WC, Grabouw
CR 12	STE-U 4253	<i>A. infectoria</i>	<i>A. infectoria</i> , clade 5	AF397229, AF404598	A, Top Red	WC, Grabouw
CR 14	STE-U 4255	<i>A. arborescens</i>	<i>Alternaria</i> sp., clade 2	AF397231, AF404599	A, Top Red	WC, Grabouw
CR 15	STE-U 4256	<i>A. infectoria</i>	<i>A. infectoria</i> , clade 5	AF397232, AF404600	A, Top Red	WC, Grabouw
CR 16	STE-U 4257	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397233, AF404601	A, Top Red	WC, Grabouw
CR 17	STE-U 4258	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397234, AF404602	A, Top Red	WC, Grabouw
CR 18	STE-U 4259	<i>A. arborescens</i>	<i>Alternaria</i> group, clade 1	AF397235, AF404603	A, Top Red	WC, Grabouw
CR 19	STE-U 4260	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397236, AF404604	A, Top Red	WC, Grabouw
CR 20	STE-U 4261	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397237, AF404605	A, Top Red	WC, Grabouw
CR 21	STE-U 4262	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397238, AF404606	A, Top Red	WC, Grabouw
CR 22	STE-U 4263	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397239, AF404607	A, Top Red	WC, Grabouw
CR 23	STE-U 4264	<i>A. arborescens</i>	<i>Alternaria</i> sp., clade 2	AF397240, AF404608	A, Top Red	WC, Grabouw
CR 24	STE-U 4265	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397241, AF404609	A, Top Red	WC, Grabouw
CR 25	STE-U 4266	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397242, AF404610	A, Top Red	WC, Grabouw
CR 26	STE-U 4267	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF397243, AF404611	A, Top Red	WC, Grabouw
CR 27	STE-U 4268	<i>A. tenuissia</i>	<i>A. tenuissia</i> , clade 4	AF397244, AF404612	A, Top Red	WC, Grabouw
CR 28	STE-U 4269	<i>A. infectoria</i>	<i>A. infectoria</i> , clade 5	AF397245, AF404613	A, Top Red	WC, Grabouw
CR 30	STE-U 4271	<i>A. infectoria</i>	<i>A. infectoria</i> , clade 5	AF397247, AF404614	A, Top Red	WC, Grabouw
CR 31	STE-U 4272 EGS 27-193 ^d	<i>A. infectoria</i>	<i>A. infectoria</i> , clade 5	AF397248, AF404615	Triticu sp.	U.K.
MA 1	STE-U 4324	<i>A. tenuissia</i>	<i>A. tenuissia</i> , clade 4	AF404642, AF404616	C, Minneola tangelo	MP, Ngonini
MA 2	STE-U 4325	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF404643, AF404617	C, Minneola tangelo	MP, Ngonini
MA 3	STE-U 4326	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404644, AF404618	C, Minneola tangelo	MP, Ngonini
MA 4	STE-U 4327	<i>A. tenuissia</i>	<i>Alternaria</i> group, clade 1	AF404645, AF404619	C, Minneola tangelo	MP, Ngonini
MA 5	STE-U 4328	<i>A. citri</i>	<i>Alternaria</i> group, clade 1	AF404646, AF404620	C, Minneola tangelo	MP, Ngonini
MA 6	STE-U 4329	<i>A. tenuissia</i>	<i>A. tenuissia</i> , clade 4	AF404647, AF404621	C, Minneola tangelo	MP, Ngonini
MA 7	STE-U 4330	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404648, AF404622	C, Minneola tangelo	MP, Ngonini
MA 8	STE-U 4331	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404649, AF404623	C, Minneola tangelo	MP, Ngonini

Table 1 cont.

Accession no.	Other no.	Morpho-species ^a	Phylospecies	Genank accession no. ITS, Histone	ariety or cultiar ^b	Location ^c
MA 9	STE-U 4332	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404650, AF404624	C, Minneola tangelo	MP, Ngonini
MA 10	STE-U 4333	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404651, AF404625	C, Minneola tangelo	MP, Ngonini
MA 11	STE-U 4334	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404652, AF404626	C, Minneola tangelo	WC, Citrusdal
MA 12	STE-U 4335	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404653, AF404627	C, Minneola tangelo	NP, Letaba
MA 13	STE-U 4336	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404654, AF404628	C, Minneola tangelo	NP, Letaba
MA 14	STE-U 4337	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404655, AF404629	C, Nael orange	WC, Citrusdal
MA 15	STE-U 4338	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404656, AF404630	C, Nael orange	WC, Citrusdal
MA 16	STE-U 4339	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404657, AF404631	C, Nael orange	WC, Citrusdal
MA 17	STE-U 4340	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404658, AF404632	C, Nael orange	WC, Citrusdal
MA 18	STE-U 4341	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404659, AF404633	C, Cleentine	WC, Citrusdal
MA 19	STE-U 4342	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404660, AF404634	C, Cleentine	WC, Citrusdal
MA 20	STE-U 4343	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404661, AF404635	C, Cleentine	WC, Citrusdal
MA 21	STE-U 4344	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404662, AF404636	C, Cleentine	WC, Citrusdal
MA 22	STE-U 4345	<i>A. arborescens</i>	<i>A. arborescens</i> , clade 3	AF404663, AF404637	C, Cleentine	WC, Citrusdal
MA 23	STE-U 4346	<i>A. alternata</i>	<i>Alternaria</i> group, clade 1	AF404664, AF404638	C, Cleentine	WC, Citrusdal
MA 24	STE-U 4347	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404665, AF404639	C, Cleentine	WC, Citrusdal
MA 25	STE-U 4348	<i>A. pellucida</i>	<i>Alternaria</i> group, clade 1	AF404666, AF404640	C, Cleentine	WC, Citrusdal
MA 26	STE-U 4349	sterile	<i>Alternaria</i> group, clade 1	AF404667, AF404641	C, Minneola tangelo	MP, Ngonini

^a based on cultural characteristics Serdani et al. 2001, and sporulation groups Sions Roberts 1993.

^b A, AppleC, citrus.

^c PrinceMP, MpualangaNP, Northern PrinceWC, Western Cape.

^d Reference isolate of *A. infectoria*.

necessary. Alignment gaps were coded as fth base in the analysis. Phylogenetic analyses were performed with PAUP version 4.0b6 (Swofford 2001). A partition homogeneity test in PAUP was conducted for the ITS and histone sequence alignment to evaluate the possibility of a joint analysis of the two data sets. The most parsimonious trees were determined from the individual and combined data sets using the heuristic search options with 1000 random sequence input orders with MULPARS on and TR branch swapping for the exact solution. The unconstrained topologies of the equally parsimonious trees were compared using the Kishino-Hasegawa test of PAUP. The best topology was selected as the most parsimonious tree topology. Parsimony bootstrap with 1000 replicates in PAUP was applied to the tree to evaluate the stability. Other measures including tree length, consistency, retention, rescaled consistency and homoplasy indices TL, CI, RI, RC and HI were also calculated. The aic likelihood analyses with 100 sequence randomisations

were also performed for the data sets to test the parsimonious tree topology. Substitution model was selected for the likelihood setting with transition/transversion ratio estimation. The distinct key sequence insertions exist in various regions in both ITS and histone data sets. In order to avoid the effect of these insertions to their local regions in the alignment, analyses excluding the insertions were conducted. As nucleotide composition of these key sequence insertions distinguished the species involved, aic likelihood analysis with 1000 sequence randomisations and substitution model of transition/transversion ratio estimation on the sub-dataset of only the key sequence insertion regions and the relevant taxa were also performed. Maximum likelihood bootstrap with 1000 replicates in PAUP was applied to the tree to evaluate the stability. Maximum parsimony analysis with exhaustive and branch and bound search options and neighbour-joining analysis with uncorrected poption were also performed for the key sequence data set. The decay indices were also

calculated using AutoDecay Eriksson 1998 to further test the robustness of the branches of all the trees generated.

RESULTS

Phylogenetic analysis

The alignment Treease accession no. SN 1195 of the joint data sets of histone and ITS spans 1116 sites in total ITS 1585, histone 5861116 including 472 constant, 428 variable but parsimony-uninformative and 216 parsimony-informative characters. Other than for minor changes, the major transition/transversion substitutions are depicted in Fig. 1 for *Alternaria arborescens*, *A. infectoria* and *A. tenuissia*.

The maximum parsimony analysis of the individual ITS and histone data sets with 1000 random sequence input orders in heuristic search produced 2 and 288 equally most parsimonious trees (MPT) respectively.

The majority-rule consensus trees were generated with 1000 random sequence input orders and 1000 bootstrap replications in heuristic search for both data sets. The ITS gene tree distinguished two clades (Fig. 2). Isolates of *A. infectoria* clustered together with strong bootstrap support 100, while a large clade assembled all the other isolates with strong bootstrap support 100. In the large clade, two isolates of *A. tenuissia* from citrus, MA1 and MA6 formed a subclade with high bootstrap support 87. The topology of the histone gene tree data not shown is identical to the joint gene tree (Fig. 3) of the ITS and histone data sets, and also has similar bootstrap values.

The result of the partition homogeneity test ($P = 0.20$), where $P < 0.05$ was significantly incongruent showing the harmonious phylogenetic evolution of ITS and histone genes in *Alternaria* spp. and suggesting a joint analysis of the two data sets. The maximum parsimony analysis of the alignment using the heuristic search option in PAUP with 1000 random sequence input orders generated 252 MPTs and a shortest tree of 805 steps. The best tree topology of the 252 MPTs was selected as the joint gene tree (Fig. 3) through the Kishino-Hasegawa likelihood test in PAUP data not shown and evaluated with 1000 parsimony bootstrap replications in a heuristic search for clade stability. The neighbour-joining analysis of the alignment with uncorrected p and 1000 bootstrap replications in PAUP produced a majority-rule consensus tree which was topologically identical to the joint gene tree (Fig. 3) and received stronger neighbour-joining bootstrap supports data not shown to the clades of the *Alternaria* group: clade 1, *Alternaria* sp. clade 2, *A. arborescens* clade 3, *A. tenuissia* clade 4 and *A. infectoria* clade 5. Maximum-likelihood analysis in heuristic search with 100 random sequence input orders and was also performed in PAUP for the data set, which produced a tree topology data not shown identical to that of the joint gene tree (Fig. 3). The joint gene tree (Fig. 3) based on

the alignment of histone and ITS data sets formed two major clades comprising 5 smaller clades. The major clade comprising clades 1 and 2 received strong bootstrap and decay indices support 98/8. The other major clade comprising clades 3-5 also received strong bootstrap and decay indices support 100/66. Clade 1 consisted of a large number of isolates morphologically identified as *A. alternata*, *A. tenuissia*, *A. citri*, *A. arborescens* and *A. pellucida* from apple and citrus with similar branch lengths and received strong parsimony bootstrap and decay indices supports 86/2. The separation of clade 2 (CR6, CR14 and CR23) from clade 1 was strongly supported by bootstrap and decay indices 95/3. Clades 3-5 represented *A. arborescens* (CR2, CR3, CR5 and MA22), *A. tenuissia* (CR27, MA1 and MA6) and *A. infectoria* (CR12, CR15, CR28, CR30 and CR31). Each of these three clades was strongly supported by bootstrap and decay indices, namely 99/4, 99/3 and 100/7, respectively. To avoid their possible effect on local regions in the alignment, maximum parsimony, maximum-likelihood and neighbour-joining analyses with the same settings were conducted on the smaller data set excluding the key sequence insertion regions. The tree topologies data not shown generated were concordant with the joint gene tree (Fig. 3). The parsimony bootstrap values 95, 93, 97, 98, 100 for the respective clades were similar to those in the joint gene tree (Fig. 3).

Maximum parsimony analysis with exhaustive search option and midpoint rooting on the smaller data set of the key sequences of the 12 relevant taxa resulted in 4 MPTs. The maximum parsimony gene tree of the key sequences (Fig. 4) was selected using the Kishino-Hasegawa test in PAUP and evaluated with 1000 bootstrap replications in branch and bound search. The clades of *A. arborescens*, *A. tenuissia* and *A. infectoria* in the tree (Fig. 4) received strong parsimony bootstrap and decay index supports of 99/4, 100/9 and 100/38, respectively. In the *A. infectoria* clade isolates CR30 and CR31 were strongly supported by bootstrap and decay indices 99/3, indicating a core intimate genealogical tie between them. The subclade of isolates CR15 and CR28 received a bootstrap support of 81 but failed to receive decay indices. The maximum-likelihood analysis with 1000 sequence randomisations and substitution model of transition/transversion ratio estimation and the neighbour-joining analysis with uncorrected p option and 1000 bootstrap replicates were also performed for the smaller data set of the key sequences and generated tree topologies data not shown almost identical to the maximum parsimony gene tree of the key sequences (Fig. 4).

DISCUSSION

The histone and ITS nucleotide sequence gene trees obtained in the present study separated the 51 *Alternaria* isolates investigated into five clades, namely a large clade containing morphologically variable isolates,

	*65	100	587	641
MA8	GGG-----TT-ACA-----GCC-----TCC			
MA11	GGG-----TT-ACA-----GCC-----TCC			
MA20	GGG-----TT-ACA-----GCC-----TCC			
MA24	GGG-----TT-ACA-----GCC-----TCC			
MA7	GGG-----TT-ACA-----GCC-----TCC			
MA10	GGG-----TT-ACA-----GCC-----TCC			
MA12	GGG-----TT-ACA-----GCC-----TCC			
MA23	GGG-----TT-ACA-----GCC-----TCC			
MA9	GGG-----TT-ACA-----GCC-----TCC			
MA13	GGG-----TT-ACA-----GCC-----TCC			
MA14	GGG-----TT-ACA-----GCC-----TCC			
MA15	GGG-----TT-ACA-----GCC-----TCC			
MA16	GGG-----TT-ACA-----GCC-----TCC			
MA17	GGG-----TT-ACA-----GCC-----TCC			
MA18	GGG-----TT-ACA-----GCC-----TCC			
MA19	GGG-----TT-ACA-----GCC-----TCC			
MA21	GGG-----TT-ACA-----GCC-----TCC			
MA25	GGG-----TT-ACA-----GCC-----TCC			
MA2	GGG-----TT-ACA-----GCC-----TCC			
MA3	GGG-----TT-ACA-----GCC-----TCC			
MA4	GGG-----TT-ACA-----GCC-----TCC			
MA5	GGG-----TT-ACA-----GCC-----TCC			
MA26	GGG-----TT-ACA-----GCC-----TCC			
CR20	GGG-----TT-ACA-----GCC-----TCC			
CR19	GGG-----TT-ACA-----GCC-----TCC			
CR2	GGG-----TT-ACA-----GCC AGCTTTAC-TCTACAACATTACACAATCATCAGTCAC TAACAATAT--CAGGTCC			
CR7	GGG-----TT-ACA-----GCC-----TCC			
CR6	GGG-----TT-ACA-----GCC-----TCC			
CR23	GGG-----TT-ACA-----GCC-----TCC			
CR1	GGG-----T--ACA-----GCC-----TCC			
CR14	GGG-----TT-ACA-----GCC-----TCC			
CR27	GGG-----TT-ACA-----GCC AGCTTGGT---TGCAACATTACACAATCATCAGTCAGTAACAATATTTTCAGGTCC			
CR15	GGGCACTGCTTCACGGCGTGCGCGGGGGCCGGCC AGCTTCGCCTCTTCAACATTACAATAACTATCAT-ACTAACAATCTCTCAGGTCC			
CR24	GGG-----TT-ACA-----GCC-----TCC			
CR18	GGG-----TT-ACA-----GCC-----TCC			
CR8	GGG-----T--ACA-----GCC-----TCC			
CR22	GGG-----TT-ACA-----GCC-----TCC			
CR25	GGG-----TT-ACA-----GCC-----TCC			
CR30	GGGCACTGCTTCACGGCGTGCGCGGGGGCCGGCC AGCTTCGCCTCTTCAACATTACAATAACTATCAT-ACTAACAATCTCTCAGGTCC			
CR5	GGG-----TT-ACA-----GCC AGCTTTAC-TCTACAACATTACACAATCATCAGTCAC TAACAATAT--CAGGTCC			
CR31	GGGCACTGCTTCACGGCGTGCGCGGGGGCCGGCC AGCTTCGCCTCTTCAACATTACAATAACTATCAT-ACTAACAATCTCTCAGGTCC			
CR21	GGG-----TT-ACA-----GCC-----TCC			
MA1	GGG-----TT-ACA-----GCC AGCTTGGT---TGCAACATTACACAATCATCAGTCAGTAACAATATTTTCAGGTCC			
MA6	GGG-----TT-ACA-----GCC AGCTTGGT---TGCAACATTACACAATCATCAGTCAGTAACAATATTTTCAGGTCC			
MA22	GGG-----TT-ACA-----GCC AGCTTTAC-TCTACAACATTACACAATCATCAGTCAC TAACAATAT--CAGGTCC			
CR12	GGGCACTGCTTCACGGCGTGCGCGGGGGCCGGCC AGCTTCGCCTCTTCAACATTACAATAACTATCAT-ACTAACAATCTCTCAGGTCC			
CR17	GGG-----TT-ACA-----GCC-----TCC			
CR16	GGG-----TT-ACA-----GCC-----TCC			
CR3	GGG-----TT-ACA-----GCC AGCTTTAC-TCTACAACATTACACAATCATCAGTCAC TAACAATAT--CAGGTCC			
CR26	GGG-----TT-ACA-----GCC-----TCC			
CR28	GGGCACTGCTTCACGGCGTGCGCGGGGGCCGGCC AGCTTCGCCTCTTCAACATTACAATAACTATCAT-ACTAACAATCTCTCAGGTCC			
	752		821	
MA8	AAG-----AGT-----GCG-----CCG			
MA11	AAG-----AGT-----GCG-----CCG			
MA20	AAG-----AGT-----GCG-----CCG			
MA24	AAG-----AGT-----GCG-----CCG			
MA7	AAG-----AGT-----GCG-----CCG			
MA10	AAG-----AGT-----GCG-----CCG			
MA12	AAG-----AGT-----GCG-----CCG			
MA23	AAG-----AGT-----GCG-----CCG			
MA9	AAG-----AGT-----GCG-----CCG			
MA13	AAG-----AGT-----GCG-----CCG			
MA14	AAG-----AGT-----GCG-----CCG			
MA15	AAG-----AGT-----GCG-----CCG			
MA16	AAG-----AGT-----GCG-----CCG			
MA17	AAG-----AGT-----GCG-----CCG			
MA18	AAG-----AGT-----GCG-----CCG			
MA19	AAG-----AGT-----GCG-----CCG			
MA21	AAG-----AGT-----GCG-----CCG			

Fig. 1. For legend see opposite page.

the *Alternaria* group clade 1, *Alternaria* sp. clade 2, *A. arborescens* clade 3, *A. tenuissia* clade 4 and *A. infectoria* clade 5.

Alternaria black rot of citrus has traditionally been ascribed to *A. citri* rown Eckert 1988. Contrary to this belief, however, our ndings show that Sions

1999awas correct in stating that seeral sall-spored species of *Alternaria* are inoled in causing this disease. Although the ITS data set only distinguished a large *Alternaria* clade and *A. infectoria* Fig. 2, the cobined ITS and histone data set separated ore taa frothe large *Alternaria* group clade 1, naely *Alternaria* sp.

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MA25  AAG-----AGT-----GCG-----CCG
MA2   AAG-----AGT-----GCG-----CCG
MA3   AAG-----AGT-----GCG-----CCG
MA4   AAG-----AGT-----GCG-----CCG
MA5   AAG-----AGT-----GCG-----CCG
MA26  AAG-----AGT-----GCG-----CCG
CR20  AAG-----AGT-----GCG-----CCG
CR19  AAG-----AGT-----GCG-----CCG
CR2   AAGTCCGCACCGTTAGT-ACCATCTTCATTCATCTTGCGATGCATC-TTTCTAAC---ATGCTCCAG
CR7   AAG-----AGT-----GCG-----CCG
CR6   AAG-----AGT-----GCG-----CCG
CR23  AAG-----AGT-----GCG-----CCG
CR1   AAG-----AGT-----GCG-----CCG
CR14  AAG-----AGT-----GCG-----CCG
CR27  AAGTCCGCACCGTTAGT-ACCATCTCCACACTATCTTGCGATCCATCC-TTCTAAC---ATGCTCCAG
CR15  AAGTCCGCACCGTTAGTTACTATCCTCACCTCATCATGCGATGCATC---TCTAACGTTAATCTCCAG
CR24  AAG-----AGT-----GCG-----CCG
CR18  AAG-----AGT-----GCG-----CCG
CR8   AAG-----AGT-----GCG-----CCG
CR22  AAG-----AGT-----GCG-----CCG
CR25  AAG-----AGT-----GCG-----CCG
CR30  AAGTCCGCACCGTTGGT-ACTATC---ACTTCATCATGCGATGCACGC--TCTAAC---ATATTCCAG
CR5   AAGTCCGCACCGTTAGT-ACCATCTTCATTCATCTTGCGATGCATC-TTTCTAAC---ATGCTCCAG
CR31  AAGTCCGCACCGTTGGT-ACTATC---ACTTCATCATGCGATGCACGC--TCTAAC---ATATTCCAG
CR21  AAG-----AGT-----GCG-----CCG
MA1   AAGTCCGCACCGTTAGT-ACCATCTCCACACTATCTTGCGATCCATCC-TTCTAAC---ATGCTCCAG
MA6   AAGTCCGCACCGTTAGT-ACCATCTCCACACTATCTTGCGATCCATCC-TTCTAAC---ATGCTCCAG
MA22  AAGTCCGCACCGTTAGT-ACCATCTTCATTCATCTTGCGATGCATC-TTTCTAAC---ATGCTCCAG
CR12  AAGTCCGCACCGTAAG--ACTATCTCACCTGATCATGCGATGCATG--TCT--CTAACATCCTTCAG
CR17  AAG-----AGT-----GCG-----CCG
CR16  AAG-----AGT-----GCG-----CCG
CR3   AAGTCCGCACCGTTAGT-ACCATCTTCATTCATCTTGCGATGCATC-TTTCTAAC---ATGCTCCAG
CR26  AAG-----AGT-----GCG-----CCG
CR28  AAGTCCGCACCGTTAGTTACTATCCTCACCTCATCATGCGATGCATC---TCTAACGTTAATCTCCAG
* positions in the alignment Mxxx

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Fig. 1. Key sequences in histone and ITS rRNA genes which can be used to design species specific probes and primers to distinguish *Alternaria arborescens*, *A. tenuissia* and *A. infectoria*.

clade 2, *A. arborescens* clade 3, *A. tenuissia* clade 4 and *A. infectoria* clade 5. The sae phenoenon was also recently obsered in a study on DCR of apple, where two separate data sets ITS seunce data, as well as secondary etabolite, pathogenicity and sporulation group datadistinguished the *A. arborescens*, *A. infectoria* and *A. tenuissia* species-groups Serdani et al. 2002. In contrast to the low resolution power of the ITS region for differentiating *Alternaria* spp., the histone data set was able to distinguish ore taa Figs 23. Roberts, Reyond Andersen 2000also concluded that the ITS region of the rDNA gene was too consered to be used as sole easure of the ariation present aong the sall-spored species of *Alternaria*. They furtherore concluded that although any of the sall-spored species of *Alternaria* were closely related, it was unwise to accept the liited ariation depicted in the ITS region as intraspecific ariation within *A. alternata*.

The *Alternaria* species inoled in the DCR cople of apple and the AR copleof citrus hae not yet been fully characteried, and presently ost papers still refer to *A. alternata* as the cause of DCR, and *A. citri* as the cause of AR Cobrink et al. 1985a, b, rown Eckert 1988. Serdani et al. 2002reported the ost coon species-group isolated froDCR syptos to be *A. arborescens*, while the *A. tenuissia* species-group proed to be the ore iportant pathogen. However, gien that a reference isolate of *A. alternata*

EGS 34-016clustered within the *A. arborescens* clade Serdani et al. 2002, we hae opted to siply refer to this large clade of isolates as the *Alternaria* group in the present study, as isolates that we regard orphologically siilar to *A. arborescens* Sions 1999b grouped in cluster 3 based on the histone seunce data Fig. 3. Furtherore, although the present olecular data could not distinguish additional taa in the larger *Alternaria* group clade 1, we still obsered morphological and cultural differences among seeral of these isolates, but were unable to relate this to presently recognized species. Whether the isolates in clade 1 really represent additional species, or siply the range of ariation within one species, will only be reealed once ore genes are studied. ased on secondary etabolite, cultural and sporulation group data Serdani et al. 2002, it sees that this clade still contains several different species. It is clear, therefore, that although the histone gene represents ore inforatie data than the ITS region, we still cannot eplain all the orphological and cultural ariation obsered aong the isolates in clade 1 Fig. 3.

With related species that are constantly eoling, it is usually accepted that the ones containing the least changes eoled frothe ore ariable ones Aise et al. 1987, Aise 1989, Harrison 1991. The large *Alternaria* group clade 1, which includes orphologically assorted isolates corresponding with the descriptions

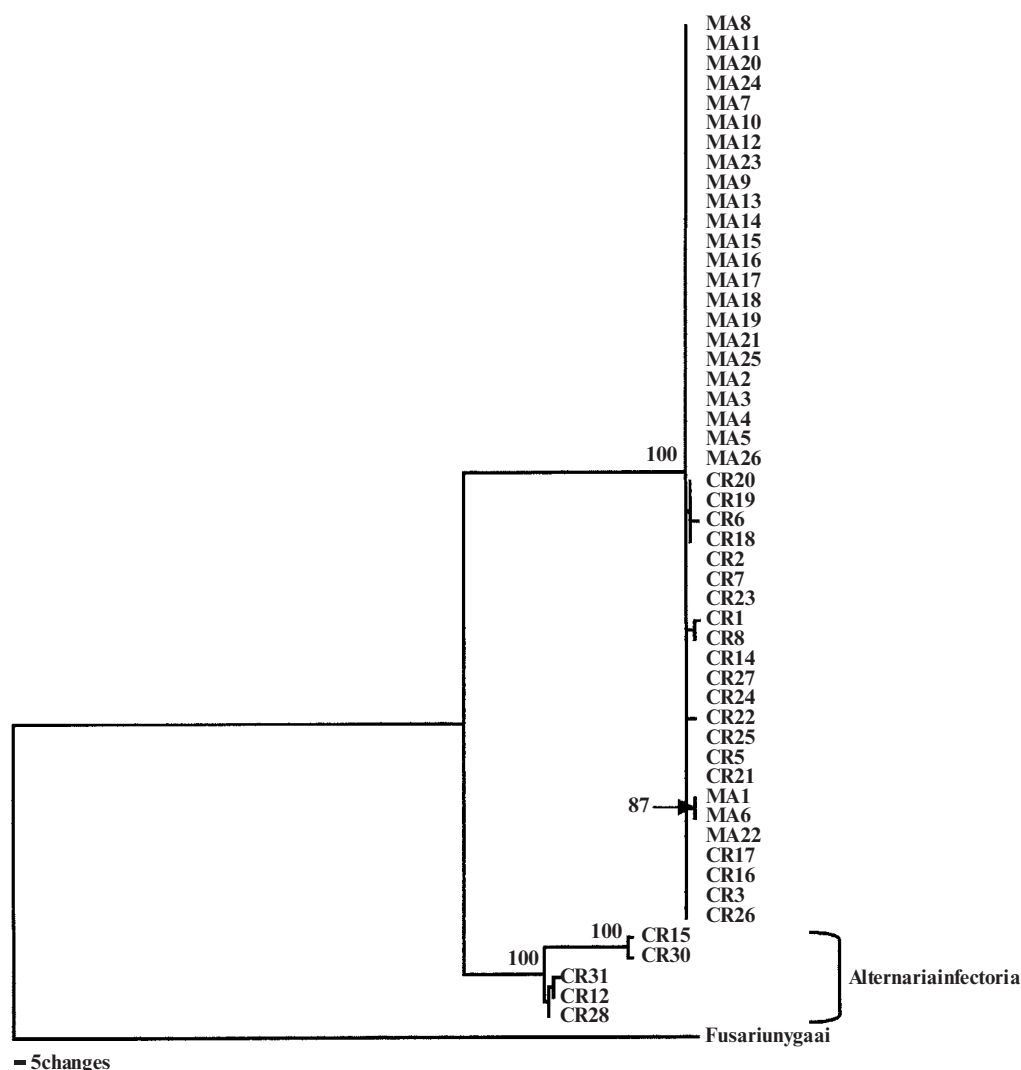


Fig. 2. One of 2 MPTs obtained from parsimony analysis of ITS using a heuristic search in the aiparsimony analysis with 1000 random sequence input orders. The tree is rooted with the outgroup *Fusariumygaai* TL 86 steps, CI 0.953, RI 0.982, RC 0.937, HI 0.047. Parsimony bootstrap values of 1000 replicates are indicated above the branches.

of *A. alternata*, *A. citri*, *A. pellucida* and *A. tenuissia*, in fact has the least substitutions in alignment. Based on ITS and histone sequence data, therefore, it can be concluded that clade 1 represents a newly evolved taxon. To the contrary, however, is the argument that the ITS region Roberts et al. 2000 as well as histone, are too conserved to distinguish all these taxa.

In the present study isolates of the *Alternaria* sp. clade 2 (CR6, CR14 and CR23) exhibited a certain number of transition/transversion substitutions. Isolates in clade 1 (*Alternaria* group) are closely related to this species clade 2. *A. arborescens* clade 3 exhibits the typical sporulation group ascribed to *A. arborescens* Sions Roberts 1993, Sions 1999b, whereas the *Alternaria* sp. (clade 2) differs in this regard. In a previous study using RAPDs, Roberts et al. 2000 delineated two groups of isolates exhibiting the *A. arborescens* sporulation pattern, thereby supporting the hypothesis that there could be more species in this complex.

The key sequences Figs 1, 4 presented here for *A. arborescens* clade 3, *A. tenuissia* clade 4 and *A. infectoria* clade 5 provide valuable references that can be used in further diagnostic studies of these pathogens. The key sequence insertion at positions 587638 for *A. arborescens*, *A. infectoria* and *A. tenuissia* proved to be unique for these taxa. A aiparsimony analysis based on a smaller data set of the key sequences of the 12 relevant taxa representing *A. arborescens*, *A. infectoria* and *A. tenuissia* (Fig. 4) separated the three clades with strong bootstrap supports of 99, 100 and 100, respectively. Furthermore, it also separated subclades within *A. infectoria*, which again suggests that additional groups exist within this species, thereby supporting the variability observed within the *A. tenuissia* species complex Sions Roberts 1993.

A further point of interest was that some species recognized to be inoculated with AR of citrus were also isolated from apples with DCR, namely the *Alternaria* group clade 1, *A. arborescens* clade 3 and

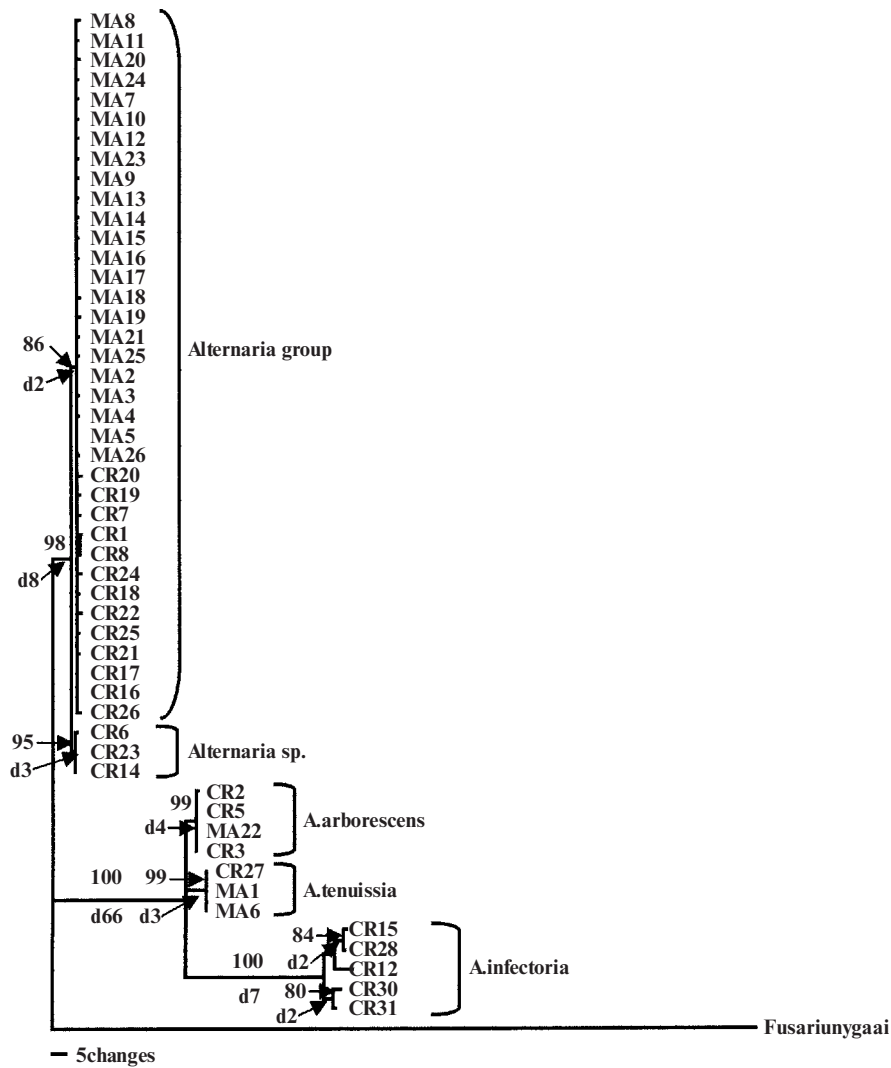


Fig. 3. One of 252 MPTs obtained from the joint alignment of ITS, and histone data sets using the heuristic search in pauparsimony with a 1000 random sequence input orders TL 805 steps, CI 0.916, RI 0.957, RC 0.876, HI 0.084. Bootstrap values obtained from 1000 replicates in heuristic search and decay indices are indicated above and below the tree branches, respectively.

A. tenuissia clade 4. Using a tooth-pick inoculation method, Serdani et al. 2002 were able to prove that *A. tenuissia* was the most important pathogen involved with DCR of apples in South Africa. Although it has not yet been determined which species plays the major role in AR of citrus, inoculation studies with representatives of the different groups identified in DCR of apple showed that the apple isolates were also pathogenic to citrus Serdani, unpubl. data. This cross-pathogenicity is further supported, therefore, by the molecular data sets obtained in the present study, where apple and citrus isolates clustered together.

Further research is still required to resolve the identity of *A. citri*, and to determine which of the CR species identified here represent the main pathogens in this disease complex. It is apparent, however, that, contrary to other reports about specificity in *Alternaria* Kusaba Tsuge 1995, Peer et al. 1999, 2000, host specificity seems to play a lesser role in post-harvest

diseases such as citrus black rot and dry core rot of apple. This is in contrast to the situation in the *Alternaria* spp. causing brown spot of citrus, where *Alternaria* isolates from rough lemon *C. abhri* and Eperor andarin *C. reticulata* caused disease on their original hosts, but not on other hosts Whiteside 1976, which in turn suggested host-specific toxins to be involved Whiteside 1976, Kohoto et al. 1979. Sions 1999b confronted the host-specific toxin producing concept employed in pathotypes of *A. alternata* Kusaba Tsuge 1994, 1995 and found that samples of these groups were actually representative of different species of *Alternaria*. In a further study, Roberts et al. 2000 concluded that, based on their pathological, morphological, biochemical and genetic data, the small-spored species of *Alternaria* represented a complex of taxa rather than a collective species. They therefore recommended that the pathotype system be abandoned. Furthermore, it was also concluded that

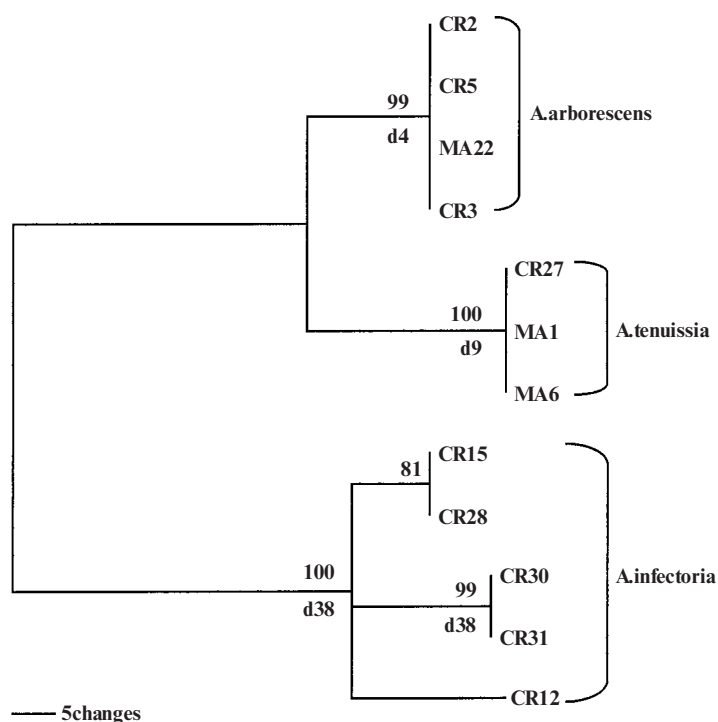


Fig. 4. One of four MPTs obtained from the key sequences of 12 selected taxa using an exhaustive search option and midpoint rooting in parsimony analysis (86 steps, CI 0.953, RI 0.982, RC 0.937, HI 0.047). Bootstrap values evaluated with 1000 bootstrap replications in branch and bound search and decay indices are shown above and below the tree branches, respectively.

A. alternata, the name commonly employed to describe a small-spored species of *Alternaria*, is rarely encountered in nature.

Masunaka et al. (2000) stated that the pathogen causing brown spot of citrus could be distinguished from the *A. citri* species based on its pathogenicity and the production of an ACT-toxin. Sions (1999a), however, segregated 135 isolates causing leaf spots on citrus into ten newly described species. All of these were morphologically distinct from both *A. alternata* and *A. citri*. A certain correlation was also seen between citrus cultivars and geographic distribution. An analysis of RAPD fragment patterns of 260 small-spored *Alternaria* isolates by Roberts et al. (2000) demonstrated that isolates from different substrates and geographic origins group together based on their three-dimensional sporulation patterns. Furthermore, their analyses also revealed that some species consisted of more than one clade or subgroup, indicating additional variation within the presently acknowledged taxa, and hence fitting with their concept of species groups containing several as yet unnamed taxa. Peeler et al. (1999) also employed RAPDs to elucidate the population structure of *Alternaria* spp. causing brown spot of citrus. An interpretation of the species concepts as proposed by Sions (1999a) led them to conclude that they could be dealing with four distinct species. However, they also found several points where their data did not agree with the concepts proposed by Sions (1999a). An isolate

of *A. lioniasperae* proved to have a RAPD profile identical to that of two reference isolates of *A. alternata*. An isolate of *A. citriarubusti* proved to be identical to two isolates of *A. citriangularis*, etc. Furthermore, several isolates associated with CR were found to cluster among isolates causing citrus brown spot, thus also questioning the differentiation between these two groups of isolates. In conclusion, Peeler et al. (1999) reported several conflicts between the morphological concepts proposed by Sions (1999a) and their molecular data sets. Roberts et al. (2000) were of the opinion that when studying small-spored *Alternaria* spp., RAPD data had to be supplemented by morphological data sets in the subsequent cluster analysis to obtain a more realistic grouping of isolates. Nevertheless, it appears that a molecular confirmation of the species associated with citrus brown spot remains lacking and this aspect will have to be resolved.

In a study focussing on the host specificity of the *Alternaria* spp. causing citrus brown spot, Peeler et al. (2000) reported *Alternaria* spp. to exist as discrete, independently evolving lineages. They further reported that the specificity observed may be due to host selection rather than tissue specificity. The only explanation for the lack of specificity observed in the present study between citrus and apple isolates is possibly due to the isolates already occurring endophytically within healthy apple and citrus tissue (Wager 1939, Serdani et al. 1998) before harvest, and that adverse climate or

cold-storage conditions stultate isolates occurring in the lower parts and calyx by giving the access to the succulent fruit tissue, resulting in rot.

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